

6-1-2008

Section: Botany, Microbiology and Zoology

## PROTECTIVE ROLE OF MELATONIN ON SOME BIOCHEMICAL ASPECTS OF NICOTINE ADMINISTRATION ON BRAIN OVARIECTOMIZED FEMALE RATS

AMIRA MERSAL

*Department of zoology, Faculty of Science,(girls) Al-Azhar University, Cairo, Egypt.*

Follow this and additional works at: <https://absb.researchcommons.org/journal>



Part of the [Life Sciences Commons](#)

---

### How to Cite This Article

MERSAL, AMIRA (2008) "PROTECTIVE ROLE OF MELATONIN ON SOME BIOCHEMICAL ASPECTS OF NICOTINE ADMINISTRATION ON BRAIN OVARIECTOMIZED FEMALE RATS," *Al-Azhar Bulletin of Science*: Vol. 19: Iss. 1, Article 2.

DOI: <https://doi.org/10.21608/absb.2008.10496>

This Original Article is brought to you for free and open access by Al-Azhar Bulletin of Science. It has been accepted for inclusion in Al-Azhar Bulletin of Science by an authorized editor of Al-Azhar Bulletin of Science. For more information, please contact [kh\\_Mekheimer@azhar.edu.eg](mailto:kh_Mekheimer@azhar.edu.eg).

---

**PROTECTIVE ROLE OF MELATONIN ON SOME BIOCHEMICAL ASPECTS OF NICOTINE ADMINISTRATION ON BRAIN OVARIECTOMIZED FEMALE RATS**

---

AMIRA TOHAMEY EBRAHIM MERSAL

*Department of zoology, Faculty of Science,(girls) Al-Azhar University.*

---

**Abstract**

The protective effect of melatonin against the damage of nicotine on the brain in ovariectomized female rats (OVX) was investigated. Cigarette smoking is common in societies worldwide and has been identified as injurious to human health. The data revealed that nicotine, a major component of cigarette smoke (at a dose of 2 mg/ KG b.wt.) resulted in a significant increase in total proteins, lipid peroxides (LPO), total cholesterol and phospholipids, and a significant decrease in acetylcholinesterase activity (AChE) and glutathione content (GSH).

Melatonin administration at dose (5 mg/kg b.wt.) to nicotine treated rats showed significantly ameliorated changes in total proteins, acetylcholinesterase (AChE), glutathione content (GSH), lipid peroxides (LPO), total cholesterol (TC) and phospholipids (Ph). The obtained results suggest that the protective effect of melatonin is mediated through decreased oxidation of lipids, thus minimizing the risk posed by nicotine.

**Introduction**

Nicotine, is one of the major hazardous components present in cigarette smoke and tobacco, plays an important role in the development of cardiovascular disease and lung cancer in smokers (Ashakumary and vijayammal, 1997; Howard *et al.*, 1998; Carpagnano, *et al.*, 2003 and Hackett *et al.*, 2003).

However, cigarette smoke has been established as a major risk factor for atherosclerosis and lung cancer (Latha *et al.*, 1992). Nicotine could play a role in atherosclerosis and contribute to acute cardiovascular events via its hemodynamic effects (Benowitz, 1997; Chan & Dimich, 2003).

In addition to its hemodynamic effects, tobacco not only has an atherogenic effect (endothelial toxicity and changes in lipid profile), but it also facilitates thrombosis and spasm (Thomas, 1993). Latha *et al.* (1993) and Ashakumary & Vijayammal, (1997) reported that nicotine administration to rats is associated with significant alterations in serum & tissue levels of lipids and lipoproteins, suggesting that nicotine may therefore, contribute at least partly to the risk posed by cigarette smoking in the development of atherosclerosis. However, the mechanisms responsible for the changes in lipids and lipoproteins are poorly understood (Criag, 1993).

In addition, lipid peroxidation is a process associated with the pathogenesis of atherosclerosis. Nicotine administration as well as cigarette smoking increased the level of lipid peroxides and the production of reactive oxygen species that may contribute to the development and/or progression of cardiovascular disease (Ashakumary and Vijayammal, 1996; Al-Senaïdy *et al.*, 1997; Helen *et al.*, 1999 and Gary & Christina, 2007).

Klevens *et al.*, (1995) discovered that cigarette smoking causes a dramatic decrease in the levels of an important enzyme that breaks down dopamine. Parkin *et al.* (1994) reported that nicotine increased Cholinergic activity which causes apoptosis (programmed cell death).

Since apoptosis helps to remove mutated or damaged cells that may eventually become cancerous, the inhibitory actions of nicotine create a more favourable environment for cancer to develop. Thus nicotine plays an indirect role in carcinogenesis. It is also important to note that its addictive properties are often the primary motivating factor for tobacco smoking, contributing to the proliferation of cancer (Parkin *et al.* 1994).

However, Zhu *et al.* (1996) noted that the majority of people diagnosed with schizophrenia smoke tobacco and for the number of schizophrenics that smoke range from 75% to 90%. It was recently argued that the increased level of smoking in schizophrenia may be due to a desire to self-medicate with nicotine.

However, Moran *et al.* (2003) reported that women smokers are more aware of their increased risk for developing lung cancer than their increased risk for developing heart disease or osteoporosis.

Waterlow *et al.*, (1978), Goldspink & Kelly, (1984). Attaix *et al.* (1988) and Sunok *et al.*, (2002) found that the rate of protein synthesis in the brain decreased with age in rats after ovariectomy. Also, Sowers (1996) investigated that not only age but also sex hormone deficiency has been shown to affect body composition and function in postmenopausal women. Estrogen increases tissue protein synthesis by stimulating transcriptional activity (Villa *et al.*, 1995 and Hofbauer *et al.*, 1999). Also, Hayase *et al.* (2001) reported that estrogen increased protein synthesis in the brain of ovariectomized female rats.

Many chemicals, drugs or smoking and radiation are known as mutagens or carcinogens. Intrinsic cellular mechanisms are engaged in cell repair, either enzymatically or via cellular reductants including glutathione and catalase which

combat oxidative damage of either intercellular metabolism or induced by extrinsic factors. Reactive free radicals are produced continuously in the body as a result of normal metabolic processes (Kehrer, 1993). extensive amounts of reactive free radicals have harmful effects that ultimately, if not removed, lead to cell damage ,mutation and cancer ( Krinsky,1989) .

Melatonin, a natural endogenous product of the pineal gland, is a highly effective antioxidant. Also, melatonin has been reported to alter the activities of enzymes which improve the total antioxidative defense capacity of the organism, i.e. glutathione peroxidase, glutathione reductase and glucose-6-phosphate dehydrogenase . (Reiter *et al.*, 1997).

Hsu *et al.* (2000) studied the protective effect of melatonin, vitamin C and beta carotene against Phosphine (an insecticide and rodenticide) – induced oxidative damage in brain, lung and liver of rats. Hsu *et al.*(2000) found that melatonin significantly or completely blocked the induced oxidative damage by phosphine while, vitamin C and beta-carotene were less effective or inactive.

Agapito *et al.* (2001) observed that oxidative damage induced by the antitumor drug, adriamycin, could be reduced by low pharmacological doses of melatonin. Moreover, vascular endothelial growth factor is the most active angiogenic factor, and the evidence of abnormally high blood levels, has been proven associated with poor prognosis in cancer patients. Melatonin was observed to control tumor growth at least in part by acting as a natural anti- angiogenic molecule (Lissoni *et al.*, 2001).

Melatonin reduced cadmium- induced lipid peroxidation in hamster brain, heart, kidney, lung and liver(Karbownik *et al.*,2001) and also prevented DNA damage induced by the pesticide, or ultraviolet radiation. Yamamoto and Mohanan, (2001) and Sener *et al.* (2002) reported that melatonin improves oxidative damage in the liver, lung and intestine induced by burn injury in rats.Furthermore, Karaoz *et al.* (2002) revealed that high doses of Vitamin C plus Vitamin E and melatonin considerably reduced chlorpyrifos-ethyle toxicity in lung tissues of rats. Exogenously administered melatonin effectively protected lungs from reperfusion injury after prolonged ischemia (Inci *et al.*, 2002). Lissoni *et al.*, (2003) suggested a new biochemotherapeutic strategy in the treatment of human neoplasms. Karaoz *et al.* (2002) found that chemotherapy was better tolerated in metastatic non-small cell lung cancer patients treated with melatonin. They also reported that melatonin modulated the effects of cancer chemotherapy, by enhancing its therapeutic efficacy and reducing its toxicity.

Accordingly, melatonin might be effective in ameliorating the progression of brain injury associated with nicotine administration. Thus, the present study was performed to investigate the possible protective effect of melatonin against nicotine-induced brain tissue injury.

### **Material And Methods**

Adult sixty female rats weighting about 120 – 140 g were used in the present study, standard diet was provided and water was available *ad libitum*. After one week of acclimatization to the laboratory environment, ten animals were served as control while other animals were ovariectomized according to the method of Agmo, (1997). Female rats were anesthetized with light diethyl ether anesthesia. With the female in a prone position, a medial dorsal incision, about 1.5 cm, long was made midway between the last rib and the knee. The skin was then pulled about 1 cm to the left and a second incision was made through the muscle layer into the peritoneal cavity. The ovaries were located through visualization of the peri-ovarian fat, prior to removal. The fat was withdrawn, The ovary was separated and the oviduct legated with silk 4.0. The ovary was cut away and the incision sutured. The ovary on the opposite side was then removed similarly through a separate incision wound closure included cut gut 4.0 for (internal) and 2.0 for (external). After two weeks from ovariectomized rats were classified into:

Group: 1: Control rat and administered distilled water

Group: 2: Ovariectomized female rats (OVX).

Group: 3: Ovariectomized+ nicotine (OVX+N).

Group: 4: Ovariectomized+ melatonine (OVX+M).

Group: 5: Ovariectomized+ nicotine + melatonine (OVX+N+M).

Nicotine was obtained from Hopkin Williams and LTD, England and was daily intraperitoneally injected at a dose of 2 mg/kg body weight for 4 weeks. Melatonin was obtained from Amoun ltd.co. Egypt in very fine powder dissolved in distilled water and given in dose of 5 mg /kg body weight for 4 weeks.

At the end of this experiment, the rats were sacrificed, the brains were isolated, weighted and homogenized for biochemical determinations according to Glowinski and Iversen ( 1966 ).Total protein content was determined by Lowry *et al.*(1951), the activity of acetylcholinesterase (AChE) in the brain was determined by the method of Gorun *et al.* (1978).

The glutathione (GSH) content in tissue homogenates was estimated by the method of Beutler (1982), the lipid peroxides (LPO) by the method of Uchiyama and Mihara (1978), total cholesterol (TC) was measured according to Allain et al. (1974), triglycerides by using the method of Wahlefeld, (1974) and phospholipids by the method of Raheja *et al.* (1973).

*Statistical analysis:-*

Data were expressed as means  $\pm$  standard error(SE) student t- test was used to elucidate the differences between treated and control group (Murray,1982). A difference was considered significant at  $P < 0.05$ .

## **Results**

Table (1) represents the brain weight and total protein content, Acetylcholinesterase activity (AChE) and glutathion content in brain tissue. The data reveals that, the ovariectomized female rats and ovariectomized treated with melatonin showed non – significant changes in the measured parameters compared to control rats.

Nicotine intraperitoneally showed a significant a increase in total proteins content, and a significant decrease in acetylcholinesterase activity (AChE) and a glutathione content (GSH) while the brain weight showed no change when compared to control. Supplementation of melatonin to ovariectomized nicotine treated rats showed a mild increase but significant decrease compared to control rats in all the measured parameters.

Table (2) represents the changes in the lipid peroxidase, triglyceride, total cholesterol and phospholipids in brain tissue of different animal groups.

Resultes in table (2) reveals that, the group of ovariectomized female rats treated with nicotine showed a significant increase in lipid peroxidase triglycerides, total cholesterol and phospholipids when compared with control group. However, ovariectomized female rats treated with nicotine and supplementation melatonin showed mild significant increase in all the measured parameters compared to control rats.

On the otherhand, the present data reveal that, the ovariectomized and the ovariectomized supplementation melatonin female rats showed non significant changes in all the measured parameter compared to control group.

**Table (1): Effect of different treatments on the (brain weight, total protein content-acetylcholinesterase activity and glutathione content) in the brain female ovariectomy rats.**

Criteria parameters	Control	OVX	OVX+N	OVX+M	OVX+N+M
Brain weight (g / tissue)	0.73±0.03	0.72±0.04	0.75±0.03	0.74±0.02	0.77±0.02
Total protein (mg/g tissue)	28.00±0.18	27.69±0.43	38.24±0.56*	27.48±0.43	25.36±0.31*
Acetylcholinesterase (μ mole/gm tissue)	11.92±0.11	11.53±0.10	9.63±0.30*	11.46±0.17	8.49±0.40*
GSH content (mg/g tissue)	0.53±0.02	0.52±0.02	0.16±0.02*	0.55±0.02	0.29±0.01*

- All values represent the mean ± S.E of 6 animals

\*Significant at P <0.05

OVX= Ovariectomized female rats.

OVX+ N= Ovariectomized female rats + Nicotine.

OVX+M= Ovariectomized female rats + Melatonin.

OVX+N+M= Ovariectomized female rats+Nicotine+Melatonin.

**Table (2): Effect of different treatments on the (lipid peroxidase level, triglyceride, total cholesterol and phospholipids) in brain female ovariectomy rats.**

Criteria parameters	Control	OVX	OVX + N	OVX + M	OVX+N+M
Lipid peroxidation nmol TBA /g tissue	187.56±0.87	187.96±0.47	219.27±0.34*	187.74±1.09	204.60±1.56*
Triglyceride (mg / g tissue)	2.18±0.10	2.78±0.04	3.28±0.04*	2.39±0.04	3.01±0.06*
Total cholesterol (mg / g tissue)	41.64±0.17	41.39±0.36	47.18±0.97*	41.19±0.66	44.66±0.58*
Phospholipid (mg / g tissue)	85.53±0.39	85.53±0.25	93.75±0.29*	85.67±0.37	89.96±0.31*

- All values represent the mean ± S.E of 6 animals

\* Significant at P < 0. 05.

OVX= Ovariectomized female rats.

OVX+N= Ovariectomized female rats+Nicotine.

OVX+M= Ovariectomized female rats+Melatonin.

OVX+N+M=Ovariectomized female rats+ Nicotine+ Melatonin.

## Discussion

The brain weight and some biochemical assay of brain ovariectomized female treated with nicotine and supplementent melation were investigated in the present study and compared to the control group.

Melatonin is a very potent and efficient endogenous free radical scavenger. It reacts with the highly toxic hydroxy radical and provides on site protection against oxidative damage to biomolecules within every cellular Compartment. Melatonin acts as a primary non–enzymatic anti oxidative defense against the devastating actions of the extremely reactive free radicals (Burkhard *et al.*, 1993; Kazez *et al.*, 2000 and Hara *et al.*, 2001). Melatonin biosynthesis has been found to be decreased in old rats (Hardeland *et al.*, 1993 and Delbarre *et al.*, 1992).

In the present study, the ovariectomized female rats with nicotine caused a significant increase in total proteins content. While supplementation of melatonin caused a significant decrease which recorded and this finding is in agreement with Touitou *et al.*,(1996) and Brzezinski, (1997). They reported discrepancy effects of exogenous melatonin and its beneficial role in biological regulation of circadian rhythms, sleep, mood, and perhaps reproduction, tumour growth and ageing.

On the otherhand, the present data showed a decrease in GSH content and this finding agreement with Change *et al.*, (1998). Gamal *et al.* (2007) reported that the reduction of tissue glutathione has been interpreted as an indicator of significant oxidant stress. Therefore, AChE activity recorded a significant decrease in the brain tissue in comparison with control rats. Finberg *et al.* (1979) reported that some drugs such as nicotine which activate post – synaptic dompamine such as L–dopa and apomorphine inhibit the release of ACh( neurotransmitter ) from brain .

In the present study the data revealed that elevation of lipid peroxidise (LPO) , triglyceride, total cholesterol and phospholipids in the brain tissues of ovariectomized female rats treated with nicotine. These results are in accordance with the observations of Allen *et al.*(1994) who reported a significant increase in total cholesterol , phospholipids and triglycerides in most of the tissues of rat and sera of nicotine treated rats. However, the changes produced in lipids fractions in nicotine administration were similar to those observed on exposure of rats to cigarette smoke (Whittaker *et al.*, 1996 and Gary& Christina, 2007), and it was felt that nicotine may therefore contribute at least partly to the risk posed by cigarette smoking in the development of atherosclerosis.



The hyperlipidemic effect of nicotine may be attributed to that nicotine increase the activity of insulin resistance leading to lipid disorders (Goldman &Klinger,1998).Nicotine administration was suggested to increase the synthesis of fatty acids in liver from carbohydrate and the mobilization of fatty acids from the depots to the liver (Lantner,1975). Tissues can synthesize fatty acids from acetyl CoA derived mainly from carbohydrates.

The main cause of impairment of acetyl CoA is intracellular carbohydrate deficiency probably induced by nicotine. Also, nicotine activates the sympathetic nervous system (by modification of action on catecholamine), where it elevated FFA, epinephrine and cortisol in the Blood of smoker (Criag, 1993).

The data of the present study are in agreement with the findings of Abd-el-Wahab (1997) who found that melatonin caused a marked decrease in parameters of hepatic damage and liver triglycerides but did not return to normal value. This protective effect of melatonin could be due to its ability to scavenge the free radical. Several authors reported that cholesterol and low density lipoprotein cholesterol (LDL-C) were reduced significantly by melatonin administration which participates in the regulation of cholesterol metabolism and in the prevention of oxidative damage to membranes Hoyos *et al.*, 2000 and Sara *et al.*, (2007). However, the melatonin administration was effective as antioxidant, although the protective effect of melatonin prevented the GSH decrease and reduced significantly the increases in enzyme activities and lipid peroxidation produced by biliary ligation (Lopez *et al.*, 2000 and Gamal *et a.*, 2007).

It could be concluded that melatonin is hypolipidemic effect of nicotine in ovariectomized female rats and may be useful in combating free radical-induced oxidative stress and tissue injury that is result of nicotine toxicity.

## References

1. Abdel- Wahab, N.M. (1997): Protective effect of melatonin against carbon tetrachloride – induced hepatic damage. *Az. J. pharm. sci.*, 19:217-224.
2. Agapito, M.T.; Antolin, Y.; Delbrijo, M.T.; Lopez-Burillo, S.and Recio, J.M. (2001): Protective effect of melatonin against Adriamycin toxicity in the rat. *J. Pineal Res.*, 31(1):23-30.
3. Agmo, A. (1997): Male rat sexual behavior. *J Brain research protocol*. 1: 203 – 209.
4. Allain, C. C., Poon, L.S., Chan, C. S., Richmond, W. and Fu, P.C. (1974): enzymatic determination of total serum cholesterol.*Clin.Chem .*, 20 : 470 – 475 .

5. Allen, S.S.; Hatsukami, D. and Gorsline, J. (1994): Cholesterol changes in smoking cessation using the transdermal nicotine system. Transdermal nicotine study group. *Prev.Med.* 23 (2): 190 – 196.
6. Al –Senaidy, A .M. Al – Zahrany, Y.A. and Al– Faqeen, M.B. (1997): Effect of smoking on serum levels of lipids peroxides and essential fat soluble antioxidants. *Nutr. Health*, 12 (1): 55 – 65.
7. Ashakumary, L.and vijayammal, P.L. (1997): Effect of nicotine on lipoprotien metabolism in rats. *Lipid*; 32 (3): 311 – 315.
8. Attaix, D. Arosseau, E.; Bayle, G.; Rosolowska, H.D. and Arnal, M. (1988): Respective influences of age and weaning on skeletal and visceral muscle protein synthesis in the lamb. *Biochem. J.*, 256: 791 – 795.
9. Benowitz, N.L. (1997): The rate of nicotine in smoking –related cardiovascular disease. *Prev. Med.*, 26 (4), 412 – 417.
10. Beutler, E. (1982): Red cell metabolism . A manual of biochemical methods 2<sup>nd</sup> edition, Publications Grune and Stratton, 89:-137-140.
11. Burkhard,P. ;Reiter, R.J.;Tan ,D.x. ;Chen,L.D. and Manchester , L.G. (1993) : Melatonin , hydroxyl radical mediated oxidative damage and aging : Ahypothesis J.pineal .Res. 14 : 151 – 168 .
12. Brzezinski, A. (1997): Melatonin in humans.*Engl.J.Med.* 336,186-195.
13. Carpagnano, G.E.; Kharitonov, S.A.; Faschino-Brbaro, M.P.; Resta, O.; Gramiccioni, E.and Barnes, P.J. (2003): Increased inflammatory markers in the exhaled breath condensate of cigarette smokers. *J. Eur.Respirology.*21 (4):589-593.
14. Chan-Yeung, M.and Dimich-Ward, H. (2003): Respiratory health effects of exposure to environmental tobacco smoke.*Respirology*, 8(2):131-139.
15. Chang, S.W.; Lanterbung, B.H. and Voehel, N.F. (1988): Endotoxin causes a neutrophil indopendant oxidant strees in rat's .*J. Appl. physiol.*, 65: 358 – 367.
16. Criag, W.Y. (1993): The effect of compounds associated with cigarette smoking on the secretion of lipoprotien in hepatic cells. *Biochem. Biophys.Act*, 1165(3): 294 – 258.
17. Delbarre, B.; Floyd, R.A.; Dellbarre, G. and Calinon, F. (1992): glutamate accumulation and decreased hydroxyl free radical. *Radical. Biol. Med.* 13: 31 – 34.
18. Finberg, J.; Buccafusco, E. and Spector, S. (1979): Regional brain acetylcholine kinetics: effect of reserpine. *Life. Sci*, 25: 147 – 156.
19. Gamal, H.; Salvatore, C.and Russel, J. (2007): Effect of chronic nicotine administration on the rat lung and liver: Beneficial role Of melatonin.*J.Sci.*19:60-67.
20. Gary, E.and Christina, N. (2007): The effect of tobacco smoke and nicotine on cognition and the brain.*J.Neurophys.*9:259-273.
21. Goldman, J. and Klinger, M. (1998): Nicotine and cardiovascular complication of chronic hypertensive disease.*Biochem, J.*, 55(2): 74 – 76.

22. Goldspink, D.F. and Kelly, F.J. (1984): Protein turnover and growth in the whole body liver and kidney of the rat from the foetus to seality. *Biochem.J.* 217:507-516.
23. Glowinski, J. and Iversen, L.L. (1966): Regional studies of catecholamines in the rat brain. I- the disposition of Norepinephrine, dopamine and dopa in various regions of the brain. *J. of Neurochemistry*, 13: 655 – 669.
24. Gorun, V.; Prionov, I.; Baitescu, V.; Balaban, G. and Barzu, O. (1978): Modified Ellman procedure for assay of cholinesterase in incude enzymatic preparations. *Anal. Biochem.* , 86(1): 324 – 326.
25. Hackett, N.R.; Heguy, A.; Harvey, B.G.; O'Connor, T.P. and Luetlich, K. (2003): Variability of antioxidant-related gene expression in the airway epithelium of cigarette smokers. *Ann.J. Respir. Cell, Mol. Biol.* 29(3):331-43.
26. Hara, M.; Yoshida, M.; Ohtani-Kaneko, R.; Shimada, A. and Hirata, K. (2001): Melatonin, a pineal secretory product with antioxidant properties, protects against cisplatin- induced nephro-Toxicity in rats. *J. Pineal Res.*, 30:129-138.
27. Hardeland, R.; Peoggeler, B.; Batzer, I. and Behrman, G. (1993): A hypothesis on the evolutionary origins of photo periodism based on circadian rhythmicity of melatonin in phylogenetically distant organisms. In: *chronobiology and chrormedicine*. Moog, eds. Peter Lang, Verlag, Frankfurt: 113-120.
28. Hayase, k.; Tanaka, M.; Tujioka, k.; Hirano, E.; Habuchi, O. And Yokogoshi, H. (2001): 17 – B – Estradiol effects brain protein synthesis rate in ovariectomized female rats. *J. Nutr.* 131: 123 – 126.
29. Kazez, A.; Demirba, M.; Ustundag, B.; Ozercan, L.H. and Saglam, M. (2000): The role of melatonin in prevention of intestinal ischemia-reperfusion injury in rats. *J. Pediatr. Surg.* 35:1444-1448.
30. Helen, A.; Rajasree, C.R.; Krishnakumar, K.; August, K.T.A and vijayammal, P.L. (1999): Antioxidant role of oils isolated from garlic and onion on nicotine induced lipid peroxidation. *Vet. Hum. Toxicol.* , 41(5): 316 – 319.
31. Hofbauer, L.C.; Khosla, S.; Dunstan; C.R.; Lacey, D.L.; Spelsberg, T.C. and Riggs, B.L. (1999): Estrogen stimulates gene expression and protein production of osteoprotegerin human osteoblastic cells. *Endocrinology*, 140: 4367 – 4370.
32. Howard, G.; Wagenknecht, L.E.; Burke, G.L.; Diezroux, A.; Evans, G.W.; MCGovern, P.; Nieto, F.J. and Tell, G.S. (1998) : Cigarette smoking and progression of atherosclerosis risk in communities (ARIC) study. *JAMA*, 279 (2): 119 – 124.
33. Hoyos, M.; Guerrero, J.M.; Perez, C.; Roilvan, J.F.; Abiani, F.; Garcia, P.A and Osuna, C. (2000): Serum cholesterol and lipid peroxidation are decreased by melatonin in diet induced hypercholesterolemic rat's. *J. Pineal. Res.*, 28 (3): 150 – 155.
34. Hsu, C.; Han, B.; Liu, M.; Yeh, C. and asida, J.E. (2000): Phosphoino -induced oxidative damage in rats: attenuation by melatonin. *Free-Radical-Biol. Med.*, 28(4):636-42.

35. Inc, I.; Inci, D.; Dutly, A.Boehler, A.andWeder, W. (2002): Melatonin Attenuates post transplanting lung ischemia-reperfusion injury. *Ann.Thorac.Surg.* 73(1):220-225.
36. Karaoz, E.; Gultekin, F.; Akdogan, M.; Oncu, M.and Gokeimen, A. (2002): Protective role of melatonin and a combination of Vitamin C and Vitamin E on lung toxicity induced by chloro -Pyriphos-ethyl in rats.*Exp.Toxicol.Pathol.* 54(2):97-108.
37. Karbownik, M.; Gitto, E.; Lewinski, A.and Reiter, R.J. (2001): Induction of lipid peroxidation in hamster organs by the carcinogen Cadmium: amelioration by melatonin.*Cell Biol. Toxicol.* 17(1):33-40.
38. Kehrer, J.P. (1993): Free radicals, a mediator of tissue injury and disease, *Crit. Rev. Toxicol.* , 23: 21 – 42.
39. Klevens, R.M.; Giovino, G.A.; Peddicord, J.P.; Nelson, D.E. ; Mowery , pand Grummer–Strawn,L.(1995): The association between veteran status and cigarette smoking behaviors.*Am.J.prev.Med* , 11 :245 – 50 .
40. Krinsky, N.I. (1989): Carotenoids and cancer in animal models, *J.Nutr.* , 119: 123 – 126.
41. Latha, M.S.; vijayammal, P.L. and kurup, P.A. (1992): Effect of nicotine on glycosaminoglycan metabolism in rats. *Indian. J.Exp. Biol*, 30(3):219– 223.
42. Latha, M.S.; vijayammal, P.L.and Kurup, P.A. (1993): Effect of nicotine administration on lipid metabolism in rats. *Indian .J. Med .Res.*, 98: 44 – 49.
43. Latner, A.B. (1975): Lipid peroxidation and antioxidants. In: *Clinical biochemistry 7<sup>th</sup> ed.*, Saunder W.B. company. Philadelphia, London, Tornoto.
44. Lissoni, P.; Chilelli, M.; Vila, S.; Ceriza, L.and Tancini, G. (2003): Five years survival in metastatic non- small cell lung cancer patients treated with chemotherapy alone or chemotherapy and Melatonin a randomized trial.*J.Pineal-Res.*, 35(1):12-15.
45. Lissoni,P.;Rovelli,F.;Malugani,F.;Bucovec,R.,Conti, A. and Maestroni, G.J. (2001): Activity of melatonin in advanced cancer patients.*Neuroendocrinology*, 22(1):45-47.
46. Lopez, P.M.; Finana, I.I.; DC, G.; UeDa, M.C and Afldmunoz, M.C. (2000); Protective effect of melatonin against oxidative strees induced by ligation of extra hepatic biliary duct in rats. Comparison with effect of L – methionine *J. Pineal. Res.*, 28 (3): 143 – 149.
47. Lowry,O.H .; Rosebrough , N.J. ; Farr, A.L.and Randall R.J.(1951): Protein measurement with the folinphenol reagent *J.Biol.chem.*, 193 : 265 – 275 .
48. Moran, S.; Glazier, G.and Armstrong, K. (2003): Women smokers'Perceptions of smoking-related health risk.*J.Women's Health (Larchmt)*, 12(4):363-71.
49. Murray, R. (1982): Scham's outline series of theory and problems of probability and statistics. McGraw – Hill book company Singapore.
50. Parkin, D.M.; Pisani, P.; Lopez, A.D. and Masuyer, E. (1994): At least one in seven cases of cancer caused by smoking. *Int. J. Cancer*, 59:494 – 504.

51. Raheja, P.K.; KAmur, C.; Singh, A. and Bhatla, I.S. (1973): New colorimetric method for the quantitative estimation of phospholipids without acid digestion .*J.lipid.Res.* 14: 695 – 697 .
52. Reiter, R.; Tang, L; Garcia, J.and JafldMounoz, H.A. (1997): pharmacological action of melatonin in oxygen radical pathophysiology.*Life.Sci*, 60(25):2255-2271
53. Sara, A.; Mahnaz, A.; Alireza, S.; Seied, R.and Bagher, M. (2007):Protective effect of melatonin on spinal cord damage after gamma irradiation.*J.Soci.for Alterna.to Animal Exper.*14:535-538.
54. Sener,G.;Sehirli,A.O.;Satiroglu,H.;Keyer-Uysal,M. and Yegan,B.(2002):Melatonin improves oxidative damage in rat model of thermal injury.*Burns*,28(5):419-425.
55. Sowers, M. (1996): Nutritional advances in osteoporosis and osteomalacia. Zeigler, M. L. filer, L. J. Ed .present knowledge in nutrition 7<sup>th</sup> ed. International life sci. Inst. 456 – 463.
56. Sunok, L.;Emi ,H ; kazyo ,T .; Yuka, M.; Kazutoshi,H . and Hidehiko, Y. (2002) : Dietary genistein affects brain protein synthesis rates in ovariectomized female rats *Am. J.soc. Nut.sci.*, 132 : 2055 – 2058 .
57. Thomas, D. (1993): Tobacco smoking and cardiovascular disease, *Respirology*.43 (10): 1218 – 1222.
58. Touitou, Y.; Selmaoui, B.; Zhao, Z.Y.; San– Martin, M. and Bogdan, A. (1996): Melatonin and biological rhythms: various aspects in human physiopathology. *Ann. pharm.*54 (6): 241 – 250.
59. Uchiyama, M. and Mihera, M. (1978): Determination of malonaldehyde precursor in tissues by thiobarbituric acid test. *Anal. Biochem.* , 86: 271 – 278.
60. Villa, E.;Camellini, L .; Dugani ,A .; Zucci, F .; Grottola , A .; Meright, A. and Manenti, F. (1995): Variant estrogen receptor messenger RNA species detected in human primary hepatocellular carcinoma. *Cancer. Res.*, 55: 498 – 500.
61. Wahlefeld, A.W. (1974): Triglycerides determination after enzymatic hydrolysis. In: methods of enzymatic analysis. (Bergmeyer, H.W, Ed) p: 1831 – 1835. Academic press, NewYork and London.
62. Waterlow,J.C;Garlick,P.J.and Millward,P.J.(1978): Protein turnover in mammalian tissues and in the whole body.*J.Nut.* 134:783-789.
63. Whittaker, P.; Wamer, W.G; Chanderbhan, R. F. and Dunkel, V.C. (1996): Effects of  $\alpha$ -tocopherol and betacarotene on hepatic lipid peroxidation and blood lipids in rats with dietary iron overload. *Nutr. Cancer*, 25 (2): 119 – 128.
64. Yamamoto, H.A. and Mohanan, P.V. (2001): Effects of melatonin on ultraviolet light exposure- induced DNA damage.*J.Pineal Res.*, 31(4):308-313.
65. Zhu, B.P.;Giovino ,G.A .;Mowery, P.D. and Eriksen , M .P. (1996) : The relationship between cigarette smoking and education revisited : implications for categorizing persons educational status . *Am. J. Public Health*, 86: 1582 –1590.

## الملخص العربي

الدور الوقائي للميلاتونين علي بعض التغيرات الكيميائية في مخ إناث الفئران  
المنزوعة المبايض والمحقونة بالنيكوتين

أميرة تهامي إبراهيم مرسال

قسم علم الحيوان - كلية العلوم - جامعة الأزهر للبنات

يهدف هذا البحث الي دراسة الدور الوقائي للميلاتونين علي بعض التغيرات الكيميائية الناتجة من حقن مادة النيكوتين علي مخ إناث الفئران والمنزوعة المبايض .

وقد قُسمت المجاميع الي خمس مجاميع وهي :

المجموعة الأولى : وهي المجموعة الضابطة وغير منزوعة المبايض

المجموعة الثانية : وهي المجموعة المنزوعة المبايض

المجموعة الثالثة : وهي مجموعة منزوعة المبايض وتم حقنها بمادة النيكوتين بجرعة 2 ميليجرام/كيلوجرام من وزن الجسم لمدة أربعة أسابيع .

المجموعة الرابعة : وهي مجموعة منزوعة المبايض وتم إعطائها مادة النيكوتين بجرعة 5 ميليجرام/كيلوجرام من وزن الجسم لمدة أربعة أسابيع .

المجموعة الخامسة : وهي مجموعة منزوعة المبايض وتم حقنها بمادة النيكوتين والميلاتونين .

وقد خلصت الدراسة الي النتائج التالية :

1 - أهدئت معاملة الفئران بالنيكوتين 2 مجم/كجم ارتفاعا ملحوظا في محتوى البروتين الكلي ومستوي الأوكسدة الفوقية للدهون والكوليستيرول الكلي والفسفوليبيدات بينما نقص نشاط انزيم الأستيل كولينستيريز ومحتوي الجلوتاثيون .

2 - عند معامل الفئران بالميلاتونين 5 مجم/كجم لوحظ تحسن ملحوظ في مستوي المعايير السابقة .

وقد خلصت الدراسة الي أن استخدام مضادات الأوكسدة ومن أهمها الميلاتونين له دور وقائي في الحماية من التعرض لأي ملوثات أو التدخين مع التقدم في العمر .